

A Practical Method for the Preparation of α' -Chloroketones of N-Carbamate Protected- α -Aminoacids

Ping Chen[†], Peter T. W. Cheng[†], Steven H. Spergel[†], Robert Zahler[†],
 Xuebao Wang[‡], John Thottathil[‡], Joel C. Barrish^{†*}, and Richard P. Polniaszek^{‡*}

Bristol-Myers Squibb Pharmaceutical Research Institute

[†]Discovery Chemistry

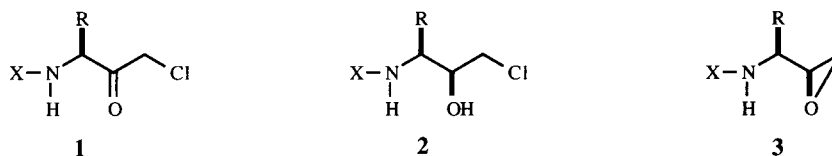
Princeton, New Jersey 08543-4000

[‡]Chemical Process Research

New Brunswick, New Jersey 08903-0191

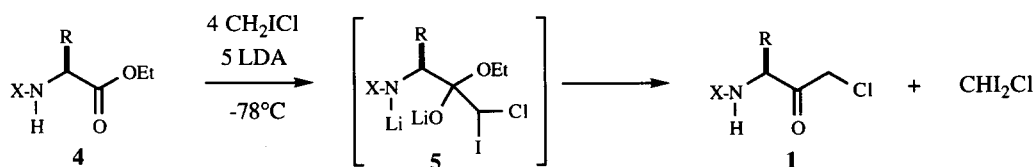
Abstract: A practical method for the preparation of α -N-BOC-epoxides from protected amino acid esters based on the Kowalski homologation reaction is described. This procedure can be readily performed on a large scale without the use of hazardous reagents and has allowed preparation of epoxides **3** in multi-kilogram quantities. © 1997 Elsevier Science Ltd.

α -N-acyl- α' -chloroketones **1** serve as irreversible enzyme inhibitors¹ and as precursors to the hydroxyethylamine isostere subunits present in many inhibitors of angiotensin converting enzyme,² renin,³ and HIV-protease.⁴ The α' -chloroketones **1** are typically converted to chlorohydrins **2**⁵ and epoxides **3**⁶ which are combined with a nucleophilic component in the preparation of the various enzyme inhibitors.



X = Boc, Cbz

The classical means of preparing α -chloroketones **1** involves conversion of an N-acyl- α -amino acid to an α -diazoketone and subsequent acidolysis with HX.⁷ Utilization of large quantities of diazomethane necessary for the production of **1** in bulk was prohibitive for practicality and safety reasons. In 1985⁸ Kowalski described a homologation of esters to α -bromoketones with the reagent system $\text{CH}_2\text{Br}_2/\text{LDA}/n\text{-BuLi}$. When this reagent system was tested on BOC-Phe-OEt, a mixture of α -bromoketone, dibromoketone and starting ester was obtained. Optimization of the reaction defined the conditions shown below.



X = Boc, Cbz

The optimum dihalomethane proved to be iodochloromethane and in this case, the $n\text{-BuLi}$ was best replaced by

excess LDA/CH₂ICl. One equivalent of LDA was necessary to deprotonate the carbamate N-H and a total of four equivalents of CH₂ICl/LDA were necessary to carry out the addition/metalation/elimination *via* intermediate **5** and drive the reaction to completion. Deprotonation of the carbamate N-H protected the chiral center from enolization and hence, racemization.⁹ Use of CH₂I₂ resulted in lower isolated yields of the iodoketone and significant amounts of a reduction product, the corresponding methyl ketone. A variety of substrates (Table) gave similar chromatographed yields of the chloroketone products. The exception was Boc-valine (**4g**) where it appears that steric hindrance prevented facile addition of the unstable chloriodomethane lithium anion, resulting in substantial recovery of starting material.¹⁰

Table

	Ester 4	1 (% Yield) ^a
4a		86
4b		77
4c		82
4d		81
4e		83
4f		79
4g		50 ^b

^a Chromatographed yield from reaction on 1 mmol scale.

^b Recovered 37% **4g**

A drawback associated with the above method is the stoichiometric production of a high boiling, toxic byproduct CH₂ICl, which made isolation problematical on scale-up. Since our ultimate goal was to prepare **3**, a

vigorously for 5 min and the lower aqueous layer discarded. Anhydrous ethanol (1500 mL) was added, the flask cooled to -78°C, and ice cold ethanolic NaBH₄ [from NaBH₄ (60 g) in anhydrous ethanol (2000 mL)] was added dropwise.

The reaction vessel was stirred at -78°C for 12h, warmed to 0°C, stirred for 2h and quenched by addition of a solution of 750 mL saturated KHSO₄ + 750 mL of water. The mixture was stirred at 0°C for 30 min, and concentrated *in vacuo*. Water (3000 mL) was added to the solid yellow residue, and the resultant mixture stirred for 30 min. The solid was collected, rinsed with water (1000 mL), hexanes (400 mL, then 600 mL, then 400 mL), and dried overnight by suction filtration affording a yellow solid, 136.1 g (76%), which analyzed by HPLC¹¹ as a 9:1 mixture of diastereomers.

The solid was taken up in hot ethyl acetate (2700 mL), cooled briefly, charcoal (8.1 g, Norit, Fisher, neutral) was added, the mixture heated briefly and filtered through celite and the celite pad washed with hot ethyl acetate (200 mL, then 50 mL). The ethyl acetate filtrate was concentrated to 1700 mL *in vacuo*, the mixture heated briefly to redissolve the precipitated chlorohydrins and allowed to cool slowly to RT, then at -5°C for 2 d. The chlorohydrin was collected as brown mossy crystals: 81.7 g, 45.5% , de 98%,¹¹ ee 99%.¹¹ The chlorohydrin **2a** was converted to epoxide **3a** with ethanolic KOH⁶ and crystallized from hexane: needles, 62.1 g, 39% overall, de 99.4%,¹¹ ee 99.9+%,¹¹ mp 121.5-123.5°C, Anal. Calcd. for C₁₅H₂₁NO₃: C, 68.42; H, 8.04; N, 5.32. Found: C, 68.32; H, 8.08; N, 5.16. Spectral data were consistent with published values.⁶

References:

- (1) Segal, D.M.; Powers, J.C.; Cohen, G.H.; Davies, D.R.; Wilcox, P.E. *Biochemistry* **1971**, *10*, 3728. Powers, J.C.; Tuhy, P.M. *Biochemistry* **1973**, *12*, 4767. Tsuda, Y.; Okada, Y.; Nagamatsu, Y.; Okamoto, U. *Chem. Pharm. Bull.* **1987**, *35*, 3576.
- (2) Gordon, E. M.; Godfrey, J. D.; Pluscec, J.; von Langen, D.; Natarajan, S. *Biochem. Biophys. Res. Commun.* **1985** *126*, 419-426.
- (3) Luly, J.R.; Plattner, J.J.; Stein, H., Yi, N.; Soderquist, J.; Marcotte, P.A.; Kleinert, H.D.; Perun, T.J. *Biochem. Biophys. Res. Comm.* **1987**, *143*, 44.
- (4) Review: Thaisrivongs, S. *Ann. Rep. Med. Chem.* **1994**, *29*, 133.
- (5) Rotella, D.R. *Tet. Lett.* **1995**, *36*, 5453. Albeck, A.; Persky, R. *Tetrahedron* **1994**, *50*, 6333.
- (6) Evans, B.E.; Rittle, K.E.; Homnick, C.F.; Springer, J.P.; Hirshfield, J.; Veber, D.F. *J. Org. Chem.* **1985**, *50*, 4615. Luly, J.R.; Dellaria, J.F.; Plattner, J.J.; Soderquist, J.L.; Yi, N. *J. Org. Chem.* **1987**, *52*, 1487. Askin, D.; Wallace, M.A.; Vacca, J.P.; Reamer, R.A.; Volante, R.P.; Shinkai, I. *J. Org. Chem.* **1992**, *57*, 2771. Barrish, J. C.; Gordon, E.; Alam, M.; Lin, P.-F.; Bisacchi, G. S.; Chen, P.; Cheng, P. T. W.; Fritz, A. W.; Greytok, J. A.; Hermsmeier, M. A.; Humphreys, W. G.; Lis, K. A.; Marella, M. A.; Merchant, Z.; Mitt, T.; Morrison, R. A.; Obermeier, M. T.; Pluscec, J.; Skoog, M.; Slusarchyk, W. A.; Spengel, S. H.; Stevenson, J. M.; Sun, C.-Q.; Sundeen, J. E.; Taunk, P.; Tino, J. A.; Warrack, B. M.; Colonno, R.; Zahler, R. *J. Med. Chem.* **1994**, *37*, 1758. Green, B.E.; Chen, X.; Norbeck, D.W.; Kempf, D.J. *Synlett* **1995**, 613. Beaulieu, P.L.; Wernic, D.; Duceppe, J.-S.; Guindon, Y. *Tet. Lett.* **1995**, *36*, 3317. Ng, J.S.; Pryzbyla, C.A.; Liu, C.; Yen, J.C.; Mueller, F.W.; Weyker, C.L. *Tetrahedron* **1995**, *51*, 6397. Heinsoo, A.; Raidaru, G.; Linask, K.; Jarv, J.; Zetterstrom, M.; Langel, U. *Tetrahedron: Asymmetry* **1995**, *6*, 2245.
- (7) Powers, J.C.; Wilcox, P.E. *J. Am. Chem. Soc.* **1970**, *92*, 1782.
- (8) Kowalski, C.J.; Serajul Haque, M. *J. Org. Chem.* **1985**, *50*, 5140. Kowalski, C.J.; Reddy, R.E. *J. Org. Chem.* **1992**, *57*, 7194.
- (9) Buckley III, T.F.; Rapoport, H. *J. Am. Chem. Soc.* **1981**, *103*, 6157.
- (10) Attempts to pre-form the anion followed by reaction with **4g** gave only recovered starting material.
- (11) Determined by HPLC analyses on C18 reverse phase (de) or chiral (ee) Daicel (or Chiralpak AD for **1a**) stationary phases. The enantiomeric purity of the chloromethylketone **1c** was also determined and found to be 98.4%. We thank Dr. W. Thompson, Mr. J. Venesky, and Ms. B. Beyer of our Analytical Research and Development Department for these analyses.
- (12) A procedure to prepare α -haloketones of N-dibenzyl protected amino acids by addition of a halomethyl lithium reagent to the corresponding esters was recently reported: Barluenga, J.; Baragana, B.; Alonso, A.; Concellon, J.M. *J. Chem. Soc., Chem. Commun.* **1994**, 969.
- (13) Process for Preparing N-Protected Amino Acid α -Halomethyl Ketones and Alcohols From N-Protected Amino Acid Esters. Barrish, J.C.; Thottathil, J.K.; Cheng, P.T.W.; Chen, P.; Spengel, S.H.; Zahler, R.; Polniaszek, R.P.; Wang, X. U.S. Patent 5481011, 1996.

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